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Assessment of Bacterial Load in the Fresh Water Lake System of Tamil Nadu

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ABSTRACT

Keywords

Freshwater, Lakes, Coliform Bacteria, Bacterial Diversity.

Article Info

Accepted: 12 May 2016 Available Online: 10 June 2016 Microbial study was done on two freshwater systems located in Viluppuram District of Tamil Nadu. Total bacterial density ranged from 6.2 - 14.4×10⁷ cfu/lit in Mukaiyur lake and from 5.4 - 8.6×10⁶ cfu/lit in Tirukoilur lake. Maximum Total Bacterial Density was noticed in November in both systems. Total coliform ranged from 165 - 1635/100 ml in Mugaiyur lake and from 315 - 521/100 ml in Thirukoilur lake. Mukaiyur lake recorded 22 bacterial species belonging to 16 genera, while Thirukoilur lake recorded 28 species belonging to 18 genera. In both the systems there were 11 bacterial perennial species occurred during study period.

Introduction

Microorganisms characterized are by tremendous metabolic versatility. Many important ecosystem processes rely entirely on microbial activities (e.g. nitrogen fixation). Moreover, viruses can cause diversification bacterial through transfers 'Weinbauer and Rassoulzadegan -2004' which in turn, has the potential to influence ecosystem functioning and the services mediated by microorganisms.

Eventhough great strides have been made in describing microbial diversity by means of modern analytical methods and a range of essential microbial-mediated processes has also been measured. Most assessments of microbial community structure and the activities that impact ecosystem functioning have been examined outside the conceptual framework of ecology and, with few exceptions, relationships between microbial biodiversity and ecosystem functioning have not been explicitly addressed.

Given the many competitive and facilitative interactions known among bacteria, changes in their biodiversity potentially have enormous scope to affect ecosystem processes (Heard and Richardson, 1995). Hence the present study was attempted to analyse the bacterial diversity in a freshwater system located in Villuppuram District, Tamil Nadu.

Materials and Methods

The freshwater systems chosen for the present study are Mugaiyur lake and Tirukoilur lake located in Villupuram District of Tamil Nadu (Plate 1). Of these one is located in Mugaiyur village and referred to as Mugaiyur lake (Plate 2). The second lake is located in Tirukoilur town and referred to as Tirukoilur lake (Plate 3). Both the lakes are located at an elevation of about 85' MSL. The distance between two lakes is about 16 km and hence the general wealth conditions are almost similar to both lakes. However, both the lakes depend on varied water sources even though both are perennial. While Mugaiyur lake is rainfed, the Tirukoilur lake is purely river fed (Penniyar river of Villupuram District).

The water samples were collected by using aseptically broad mouthed sterilized glass containers, transferred to ice boxes and immediately brought to the laboratory for culturing within 6 hours. In addition, water samples were also collected in sterilized bottles for bacterial studies and transferred immediately to the laboratory in compact thermo- cold ice boxes with minimum exposure to light, temperature and undue shaking for further analyses. Water samples for both the systems were collected between 7:30 and 8:30 am.

WHO (1985) guidelines were followed for the frequency of sampling. Total bacterial count (TBC) was estimated by standard plate count using nutrient agar, while Most Probable Number (MPN), Total Coliform (TC) and faecal *Streptococci* (FS) were carried out by multiple tube method using Mc Conkey's and sodium azide broth respectively as recommended by ISI (1982), WHO (1985) and APHA (2000). Other biochemical tests were also done for isolation and identification of different

bacterial species by using selected media and tests like IMVIC, Gram stain, fermentation, nitrate reductase, oxidase, citrate, catalase and H₂S, as recommended by Jacobs and Gerstein (1960), Sirockin and Cullimore (1969), Crueckshank *et al.* (1975) and APHA (1985).

Fluctuations of Microbiological Variables

Bacterial Abundance and Diversity (Tables 1-4)

Total Bacterial Density (TBD)

The Total Bacterial Density in the surface water of Mugaiyur lake ranged from 6.2 to 14.4×10^7 cfu/l and for the Tirukoilur lake from 5.4 to 8.6×10^6 cfu/l. For the bottom water, Mugaiyur lake recorded levels ranging from 6.4 to 14.8×107 cfu/l and for the Tirukoilur lake from 6.0 to 8.8×10^7 cfu/l. The bottom water TBD of both systems followed the same pattern as that of surface water recording minimum levels in January and maximum in November. However a comparison of TBD levels reveals that bottom water recorded higher levels than surface water in both the systems (Tables 1 and 2). Similar observations were also noticed by Shimna (2012), Sankar Rao (2013) and Mugilan (2014). The maximum TBD noticed during November in present study is due to the rains resulting in the mixing of waters along with run-off water entering the systems bringing nutrients for enabling the growth of the organisms while the minimal values noticed in January may be due to the lower water temperature which is unsuitable for the growth of bacteria. Similar observations were also recorded by Shimna (2012), Sankarrao (2013) and Mugilan (2014). A perusal of concentrations of Oxidable Organic Mater, Ammonia nitrogen, Nitrate-nitrogen and Phosphate (OOM, NH₃-N, NO₃-N and PO₄) levels in the waters of both the systems reveals a higher concentration during the rainy season which could also be a reason for the highest TBC shown during this season. In the present study, correlation between TBC and oxidizable organic matter, suspended solids and ammonia showed a positive relationship indicating their effect on bacteria. The differences in bacterial count noticed during the different seasons of the year can also be attributed to the differences in temperature and suspended solid levels as suggested by many workers (Seki, 1972; Fukami et al., 1983; Sivakami et al., 2011, 2012) in addition to oxidizable organic matter and nutrients (Sivakami et al., 2012; Sankarrao, 2013).

A comparison of the nutrient levels in different seasons also reveal that the TBC was high when the nutrient levels were also high. Correlation between TBC nutrients like phosphates, nitrates sulphates also a revealed positive correlation in both the systems. However, Jones (1971), suggested that temperature, pH and oxygen are the main limiting factors controlling the vertical bacterial population. A comparison of TBC among the two systems reveals that Mugaiyur lake recorded higher levels than Tirukoilur lake. This is probably due to higher amount suspended of solids. oxidizable organic matter as well as nutrients in this system.

Total Coliform Count

Total Coliform Count (TCC) in the surface water of Mugaiyur lake was found to range from 165 to 1635 per 100 ml and from 315 to 521 per 100 ml in the Tirukoilur lake. The bottom water TCC levels in Mugaiyur lake ranged from 205 to 1685 per 100 ml and from 330 to 585 per 100 ml in Tirukoilur lake. Both the surface and bottom waters in both the systems followed the same pattern

by recording minimal levels in January and the maximum in November (Tables 1 & 2).

In the present study, the maximum TCC that was recorded in November in both systems can again be attributed to entry of water into the system due to rains resulting in surface run-off entering the systems resulting in suspended solids, OOM and nutrients. Correlating TCC with suspended solids, OOM and nutrients (NO₃, NH₃, SO₄, PO₄) reveals a positive correlation in both the systems. The higher TCC levels noticed in the bottom waters when compared to surface waters can again be attributed to increased amount of Suspended Solids and Oxidable Organic Matter (SS and OOM) and nutrients in the bottom water when compared to the surface water.

Among the two systems, Mugaiyur lake recorded a higher level when compared to that of the Tirukoilur lake which may be due to higher amount of suspended solids, temperature, OOM and nutrients like PO₄, NO₃ and NH₃. Literature reveals that similar findings were also suggested by Kumar and Shaka (2009), Petter *et al.* (2012), Shimna (2012), Sankar Rao (2013) and Mugilan (2014).

Faecal Streptococci (FS)

Faecal *Streptococci* in the surface water of Mugaiyur lake was found to range from 82 to 482 per 100 ml and from 102 to 252 per 100 ml in the Tirukoilur lake. The bottom water Faecal *Streptococci* count in Mugaiyur lake was found to range from 92 to 562 per 100 ml and from 132 to 257 per 100 ml in Tirukoilur lake. The bottom water Faecal *Streptococci* count followed the same pattern of surface waters in both the systems recording minimum level in January and maximum in November (Tables 1 and 2).

 Table.1 Bacteriological Studies in Mugaiyur Lake Water

	Tota	al Bacterial (per 1	Density (T 00 ml)	(BD)	Tota		s Count (T 00 ml)	CC)	Faecal Streptococci Count (FSC) (per 100 ml)					
	Surface water		Botton	n water	Surface	e water	Botton	ı water	Surface	e water	Botton	n water		
Month	2013	2014	2013	2014	2013	2014	2013	2014	2013	2014	2013	2014		
January	6.2	6.7	6.4	6.9	165	225	205	295	112	82	142	92		
February	6.7	6.9	6.7	7.0	205	235	335	315	142	102	152	122		
March	7.6	7.6	7.8	7.8	305	345	385	375	162	142	172	161		
April	7.8	7.8	7.9	8.0	345	375	410	405	170	151	182	182		
May	8.1	8.2	8.0	8.4	375	435	485	445	172	162	192	192		
June	8.4	8.7	8.2	8.9	390	485	505	515	177	172	205	192		
July	9.1	8.8	8.4	9.0	410	510	555	585	181	182	220	212		
August	10.6	9.0	10.8	9.1	465	535	605	665	184	197	246	231		
September	10.8	9.2	11.0	9.8	605	585	635	735	242	242	262	252		
October	13.2	12.9	13.4	13.2	805	735	825	985	312	302	352	332		
November	14.4	13.6	14.8	13.9	1445	1635	1805	1685	482	442	562	562		
December	11.4	11.2	11.9	11.8	865	945	925	1035	362	352	402	392		

 Table.2 Bacteriological Studies in Tirukoilur Lake

	Tota	al Bacterial (per 1	Density (T 00 ml)	(BD)	Tot		s Count (To	CC)	Faecal Streptococci Count (FSC) (per 100 ml)					
	Surfac	e water	Botton	n water	Surfac	e water	Bottom	n water	Surface	e water	Botton	n water		
Month	2013	2014	2013	2014	2013	2014	2013	2014	2013	2014	2013	2014		
January	5.4	6.0	6.0	6.6	315	325	330	336	102	109	122	141		
February	5.8	6.2	6.2	6.8	345	341	350	371	107	117	127	132		
March	5.9	6.8	6.4	7.0	360	365	366	375	109	119	132	137		
April	6.1	6.9	6.5	7.2	385	381	390	396	112	123	134	147		
May	6.4	7.1	6.7	7.4	390	396	401	405	123	128	139	152		
June	6.7	7.3	7.1	7.5	410	401	422	411	142	132	147	157		
July	6.8	7.5	7.4	7.7	415	422	430	455	172	155	177	187		
August	7.0	7.6	8.0	7.9	445	449	460	471	192	162	197	192		
September	7.1	7.8	8.1	8.2	465	482	490	495	202	192	217	212		
October	7.4	8.1	8.3	8.4	475	498	516	524	212	222	227	237		
November	8.6	8.2	8.8	8.5	490	521	584	585	232	252	247	257		
December	7.2	7.8	7.4	7.5	415	435	425	465	176	182	217	207		

 Table.3 Seasonal Changes in Bacterial Composition of Mugaiyur Lake

No.	Species	Year	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec
1.	Aerobacter aerogenes	2013	+	+	+	+	+	+	+	+	+	++	+++	+++
		2014	+	+	+	+	+	+	+	+	+	++	+++	+++
2.	Aeromonas hydrophila	2013	+	+	+	+	+	+	+	+	+	++	++	++
		2014	+	+	+	+	+	+	+	+	+	++	++	++
3.	Bacillus cereus	2013	+	+	+	_	_	_	+	+	+	++	++	++
		2014	+	+	+	_	_	_	+	+	+	++	++	++
4.	Bacillus firmus	2013	++	++	++	++	++	++	++	++	+++	+++	+++	+++
		2014	++	+	++	++	+++	++	++	++	++	+++	+++	++++
5.	Bacillus subtilis	2013	+++	+++	+++	+++	++	++	++	++	+++	++++	++++	++++
		2014	+++	+++	++	+++	+++	++	+	++	++	++++	++++	++++
6.	Clostridium perfringens	2013	+	_	_	_	_	+	+	+	+	+	++	++
		2014	+	_	_	_	_	_	+	+	+	+	++	++
7.	Enterobacter aerogenes	2013	+	+	+	+	+	+	+	+	++	++	++	+
	_	2014	+	+	+	+	+	+	+	+	++	++	++	+
8.	Enterobacter cloacae	2013	+	+	+	_	_	_	_	+	+	+	+	+
		2014	+	+	+	_	_	_	_	_	+	+	+	+
9.	Erwinia rhapontici	2013	+	_	_	_	_	_	_	_	_	+	+	+
		2014	_	_	_	_	_	_	_	_	_	_	_	_
10.	Escherichia coli	2013	++++	++++	+++	+++	++++	+++	+++	+++	++++	++++	+++++	+++++
		2014	++++	++++	+++	+++	++++	+++	+++	+++	++++	++++	++++	++++
11.	Flavobacterium johnsoniae	2013	+++	+++	+++	+++	+++	++	++	++	+++	+++	+++	+++
		2014	+++	++	++	++	++	++	++	+++	++	+++	+++	+++
12.	Klebsiella pneumoniae	2013	+	+	+	_	_	+	+	+	++	++	++	+
	-	2014	+	+	+	_	_	+	+	+	++	++	++	++
13.	Proteus mirabilis	2013	+++	+++	+	++	++	++	++	++	++	++	+++	+++
		2014	++	++	+	+	+	+	+	+	+	+	+++	+++
14.	Proteus vulgaris	2013	+	+	+	+	+	_	_	+	+	+	+++	+++
		2014	+	+	+	+	+	_	_	_	_	+	+++	+++

Table-3 continued....

No.	Species	Year	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec
15.	Pseudomonas aeruginosa	2013	+++	+++	++	++	++	++	++	++	+++	++++	++++	++++
		2014	+++	+++	++	++	++	++	++	++	+++	++++	++++	++++
16.	Salmonella typhi	2013	+	_	_	_	_	_	+	+	+	+	+	+
		2014	+	_	_	_	_	_	_	+	+	+	+	+
17.	Serratia marcescens	2013	+	+	+	+	_	_	_	_	_	+	+	+
		2014	+	+	+	+	_	_	_	_	_	+	+	+
18.	Shigella sonnei	2013	_	_	_	_	_	_	_	_	_	_	_	_
		2014	_	_	_	_	_	_	_	_	+	+	+	+
19.	Staphylococcus aureus	2013	++	++	++	++	+	++	++	+++	+++	+++	++++	++++
		2014	++	++	++	++	+	++	++	+++	+++	+++	+++	++++
20.	Streptococcus faecalis	2013	+	+	+	+	+	+	+	++	++	++	++	++
		2014	+	+	+	+	+	+	+	++	++	++	++	++
21.	Vibrio alginolyticus	2013	_	_	_	_	_	_	_	_	_	_	_	_
		2014	+	+	_	_	_	_	_	_	_	+	+	+
22.	Vibrio cholerae	2013	+	+	+	_	_	_	+	+	+	+	+	+
		2014	+	+	_	_	_	_	+	+	+	+	+	+

^{&#}x27;+' represents Presence
'-' represents Absence

 Table.4 Seasonal Changes in Bacterial Composition of Tirukoilur Lake

No.	Species	Year	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec
1.	Aerobacter aerogenes	2013	++	++	+	+	+	+	+	+	+	+	+	+
		2014	++	++	+	+	+	+	+	+	+	+	+	+
2.	Aeromonas hydrophila	2013	+++	+++	+++	++	++	++	++	+++	+++	+++	++++	++++
		2014	+++	+++	+++	++	++	++	++	++	+++	+++	+++	++++
3.	Alcaligenes denitrificans	2013	+	+	_	_	_	_	_	_	++	++	++	++
		2014	_	_	_	_	_	_	_	_	_	+	+	+
4.	Bacillus cereus	2013	+	+	+	+	+	+	+	+	+	+	+	+
		2014	+	+	+	+	+	+	+	+	+	+	+	+
5.	Bacillus megaterium	2013	+	++	++	++	+	++	+	++	++	++	++	++
		2014	+	++	++	+	+	++	+	++	++	++	++	++
6.	Bacillus subtilis	2013	++	++	++	++	++	++	++	+++	+++	+++	+++	+++
		2014	++	++	++	++	++	++	++	++	+++	+++	+++	+++
7.	Chromobacterium sp.	2013	+	+	_	_	_	_	_	_	_	+	+	+
	_	2014	+	+	_	_	_	_	_	_	_	+	+	+
8.	Clostridium perfringens	2013	+	+	_	_	_	_	_	+	+	+	+	+
		2014	+	+	_	_	_	_	_	+	+	+	+	+
9.	Enterobacter aerogenes	2013	++	++	++	_	_	_	_	++	++	++	++	++
		2014	++	++	++	_	_	_	_	++	++	++	++	++
10.	Erwinia rhapontici	2013	_	_	_	_	_	_	_	_	+	+	+	+
	_	2014	_	_	_	_	_	_	_	_	+	+	+	+
11.	Escherichia coli	2013	+++	+++	+++	+++	+++	+++	+++	+++	++++	++++	+++	+++
		2014	+++	+++	+++	+++	++	++	++	+++	++++	++++	+++	+++
12.	Flavobacterium johnsoniae	2013	+	+	+	+	+	+	+	+	+	+	+	+
		2014	+	+	+	+	+	+	+	+	+	+	+	+
13.	Klebsiella pneumoniae	2013	+	+	+	+	+	+	+	+	+	+	+	+
	_	2014	+	+	+	+	+	+	+	+	+	+	+	+
14.	Proteus mirabilis	2013	+	+	+	_	_	_	_	_	_	+	+	+
		2014	+	+	_	_	_	_	_	_	_	+	+	+

Table-4 continued....

No.	Species	Year	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec
15.	Proteus vulgaris	2013	+	++	++	++	+	++	++	+++	+++	+++	+++	+++
		2014	+	++	++	++	+	++	++	+++	+++	+++	+++	+++
16.	Pseudomonas aeruginosa	2013	++++	++++	++++	+++	++	++	+++	+++	++++	++++	++++	++++
		2014	++++	+++	+++	+++	++	++	+++	+++	++++	++++	++++	++++
17.	Pseudomonas agarolyticus	2013	+	+	+	+	_	+	+	+	+	+	+	+
		2014	_	+	+	+	_	+	+	_	+	+	+	+
18.	Salmonella typhi	2013	+	_	+	+	_	_	_	+	+	+	+	+
		2014	+	+	+	+	_	_	_	_	+	+	+	+
19.	Serratia marcescens	2013	_	_	_	+	+	+	_	+	+	+	_	_
		2014	+	+	+	_	_	_	_	+	+	+	+	+
20.	Shigella flexneri	2013	_	_	_	_	_	_	_	_	-	_	_	_
		2014	+	+	+	+	+	_	_	_	++	++	++	++
21.	Shigella sonnei	2013	+	+	+	+	+	_	_	_	++	+++	+++	++
		2014	+	+	+	+	+	_	_	_	++	+++	+++	++
22.	Staphylococcus aureus	2013	+	+	+	+	+	+	+	+	++	++	++	++
		2014	+	+	+	+	+	+	+	+	++	++	++	++
23.	Streptococcus bovis	2013	++	++	++	+	+	+	+	+	++	+++	+++	+++
		2014	++	++	++	+	+	+	+	+	++	+++	+++	+++
24.	Streptococcus equinus	2013	+	+	+	_	_	_	_	+	+	+	++	++
		2014	+	+	+	_	_	_	_	+	+	+	++	++
25.	Streptococcus faecalis	2013	++	++	++	+	_	_	+	++	++	++	++	++
		2014	++	+	++	+	_	_	+	++	++	++	++	++
26.	Vibrio alginolyticus	2013	+	+	+	_	_	_	_	_	_	++	++	++
		2014	+	+	_	_	_	_	_	_	_	++	++	++
27.	Vibrio cholerae	2013	_	_	+	_	+	_	_	_	_	++	++	++
		2014	_	_	+	_	+	_	_	_	_	++	++	++
28.	Vibrio parahaemolyticus	2013	+	_	_	_	_	_	+	+	+	+	+	+
		2014	+	_	_	_	_	_	+	+	+	+	+	+

^{&#}x27;+' represents Presence; '-' represents Nil

Vasconcelos and Swartz (1976) reported that viability of faecal and coliform bacteria is inversely proportional to temperature, since high temperature enhances the growth of such bacteria in water. In the present study also both the systems recorded the maximum counts during rainy season (November) when temperature was low. Statistical correlation of Faecal Streptococci, total coliform count and total bacterial count temperature showed a with negative correlation (Annexures 1 to 10) thus showing their interrelationship.

Bacterial Species Diversity

Tables 3 and 4 reveal in Mugaiyur lake a total of 22 species were noticed which belonged to 16 genera while in the Tirukoilur lake, a total of 28 species belonging to 18 genera were recorded with both the systems recording 11 perennial species each. Among the perennial species, seven species (E. coli, S. aureus, B. subtilis, A. aerogenes, F. johnsomae, P. aeruginosa and A. hydrophila) were common to both the systems while one species was unique (Enterobacter cloacae) to Mugaiyur lake while for Tirukoilur lake five were unique denifrificans, (Alcaligenes megaterium, Shigella flexneri, Streptococcus bovis and S. equines). Further, in both the systems E. coli and P. aeruginosa were the dominant species in terms of count even though in Mugaiyur lake F. johnsomae also dominated while in Tirukoilur lake, A. hydrophila also dominated. With regard to the least dominant species. Mugaiyur lake recorded S. sonneri and V. alginolyticus while in Tirukoilur lake it was E. rhapontici and E. nurabilis (Tables 3 and 4).

With regard to their most favourable period of occurrence in maximal numbers, total bacterial count as a whole in both the systems recorded higher levels during the period between October and December even though each species appeared to occur in highest counts in different months of the year. Thus both the systems showed both uniqueness and similarity between them. Between the two systems, Tirukoilur lake recorded higher diversity in terms of both species as well as genera. This variation may be due to the differences in nutrient levels as reported by other workers (Lim and Flint, 1989; Bogosian *et al.*, 1996; Shimna, 2012; Sankar Rao, 2013; Mugilan, 2014).

In the present study, the presence of *Vibrio*, Aeromonas, Ε. coli, Enterobacter, Staphylococcus species etc. indicates the presence of significant levels of microbial pollution. The presence of E. coli in both the possibly indicates systems contamination. According to Kataria and Ambhore (2012) the presence of E. coli potentially indicates dangerous requiring immediate contamination attention. Hence this requires immediate attention if such systems are used for the betterment of man.

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